Forum Mini Review

The Role of Thioredoxin in the Aging Process: Involvement of Oxidative Stress

TORU YOSHIDA,¹ SHIN-ICHI OKA,² HIROSHI MASUTANI,¹,² HAJIME NAKAMURA,¹ and JUNJI YODOI¹,²

ABSTRACT

Reactive oxygen species are produced by various stressors derived from internal and external sources, including endogenous metabolic activities. Glucose metabolism is one of the most primitive sources for energy production for most cells; however, it may at the same time yield hazardous oxidative stress via simultaneous oxidant production. The protective mechanism against oxidative stress is thus an indispensable biological function. Recently, genetic mutation loci affecting life span were isolated from experimental model organisms, and several locus products were found to be closely linked with machinery either producing or defending oxidative stress. Thioredoxin (TRX) is a small protein having strong antioxiradical quenching capabilities and other multiple functions depending on the cellular redox state. In this review, we focus on the role of TRX in the aging process (senescence) as a redox-regulating molecule against oxidative stress. We also discuss the possibility of the TRX system serving as an index marker for cellular proliferation and senescence. *Antioxid. Redox Signal.* 5, 563–570.

THIOREDOXIN (TRX) AS AN ANTIOXIDANT DEPENDING ON CELLULAR REDOX STATE

CLUCOSE METABOLISM is one of the major biological systems for producing necessary ATP molecules in conjunction with mitochondrial aerobic respiration. However, this energy production system at the same time yields hazardous materials such as reactive oxygen species (ROS) and other free radical species such as nitric oxide. Active aerobic respiration may increase the chance of electrons leaking from the mitochondrial electron transfer system, which causes severe cellular damage. A protective mechanism against oxidative stress is thus an indispensable biological function.

There are many proteins and low-molecular-weight antioxidants in the typical antioxidant system. One kind is radical

quenching or scavenging proteins that include superoxide dismutase (SOD), catalase, glutathione peroxidase, glutathione reductase, TRX, peroxiredoxin, and TRX reductase. Another kind includes low-molecular-weight antioxidants such as glutathione, vitamin C, vitamin E, uric acid, and bilirubin. A small thiol protein, TRX, has recently attracted much attention due to its strong antioxiradical quenching capabilities and other important biological functions related to the regulations of cellular redox state.

TRX has been cloned as an adult T-cell leukemia-derived factor, produced by human T-cell leukemia virus-I-transformed T-cells (58, 69), or as an interleukin-1-like autocrine growth factor from Epstein–Barr virus-transformed cells (65). TRX is a 12-kDa thiol-mediated protein with a redox-active disulfide/dithiol group within the conserved active-site sequence Cys-Gly-Pro-Cys. Reduced TRX catalyzes the reduction of disulfide bonds in many proteins, and oxidized TRX is re-

¹Department of Biological Responses, Institute for Virus Research, Kyoto University, 53 Shogoin, Kawahara-cho, Sakyo-ku, Kyoto, 606-8507, Japan.

²Biomedical Special Research Unit, Human Stress Signal Research Center, National Institute of Advanced Industrial Science and Technology (AIST), Midorigaoka, Ikeda, Osaka 563-8577, Japan.

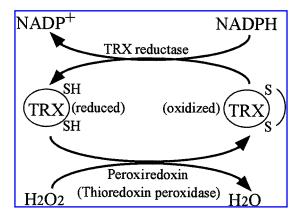


FIG. 1. TRX reducing cycle. TRX exists in either a reduced form containing two thiol groups or an oxidized form containing a disulfide bond. Oxidized TRX is reduced by TRX reductase and NADPH. The reduced form of TRX can reduce disulfide bonds of target proteins.

versibly reduced by the action of TRX reductase and NADPH (18) (Fig. 1). The TRX system (TRX, TRX reductase, and NAPDH) is widely conserved from fungus to higher eukaryotes. Several TRX-related molecules have been identified and are actively involved in the cellular antioxidant system and redox regulation with other TRX family proteins. Table 1 summarizes the TRX superfamily, which holds similar active site sequences regardless of the localization they express.

EXTENDED LIFE SPAN MUTANTS AND RESISTANCE TO OXIDATIVE STRESS

Caloric restriction (CR) has been identified as the only unequivocally accepted scheme to extend life span significantly for most organisms. Recently, the molecular basis of CR has been partially revealed as a reduced load of growth hormone signal transduction and the insulin pathway. Insulin is a primitive mitogenic factor that promotes glucose uptake by cells when the extracellular glucose concentration is high. Glucose metabolism is one of the most primitive sources for energy production for most cells, and is critical to support general biological activities. There is accumulating evidence that defective growth hormone components and insulin pathways frequently affect life span (Table 2). Since the first extended life span mutant age-1, a homologue of phosphatidylinositol 3kinase (PI3K) in mammals, was reported in C. elegans (25), several similar mutations have been identified as extending life span significantly, including daf-2 (insulin/IGF-I receptor homologue), daf-16 (forkhead transcription factor) in C. elegans (29, 34), and chico (IRS, insulin receptor substrate), InR (insulin receptor) in D. melanogaster (8, 61). In mammals, prop1df/df (Ames dwarf) and pit1dw/dw (Snell dwarf) mice both display impaired pituitary gland development and lower levels of growth hormone (3, 14), and newly reported knockout mice of insulin receptor (2) and IGF-1 receptor (19) were all found to have significantly extended life span.

Many studies have indicated that CR treatment leads to strong resistance against various oxidative stresses in many animals (1, 13). It should be also noted that extended life span mutants caused by growth hormone defects and the insulin pathway simultaneously demonstrated strong resistance to various stresses, including free radical species. Age-1 and daf-2 showed strong resistance against oxidative stress (21, 33, 55) and increased mitochondrial Mn-SOD activities (20). Moreover, many extended and reduced life span mutation genes in C. elegans have been found to be components of the mitochondrial electron transport system such as clk-1 (CoQ enzyme), isp-1 (complex III), gas-I (complex I), and mev-1 (complex II) (12, 23, 28, 32). Methuselah (mth) was first isolated as an extended life span mutant in D. melanogaster and found to show strong resistance to paraquat-induced oxidative stress (35). Several transgenic fruit flies introducing SOD and catalase (49), msra (52), and human SOD-1 (50) showed significant extension of life span. It is therefore possible that lowering the efficiency of the insulin pathway may affect energy production in mitochondria. When efficient energy production drops due to insufficient amounts of glucose,

TABLE 1. LIST OF TRX SUPERFAMILY GENES

TRX family gene	kDa	Localization	Active-site sequence
Thioredoxin (TRX)	12	Cytosol	-Cys-Gly-Pro-Cys-
Thioredoxin-2(TRX-2)	12	Mt	-Cys-Gly-Pro-Cys-
TRX-related protein (TRP32)	32	Cytosol	-Cys-Gly-Pro-Cys-
Transmembrane TRX-related protein (TMX)	30	ER	-Cys-Pro-Ala-Cys-
Sperm-specific TRX	53	Cytosol	-Cys-Gly-Pro-Cys-
Glutaredoxin (GRX)	12	Cytosol	-Cys-Gly-Tyr-Cys-
Glutaredoxin-2(GRX-2)	18	Nucleus, Mt	-Cys-Ser-Tyr-Cys-
Nucleoredoxin	48	Nucleus	-Cys-Pro-Pro-Cys-
Protein disulfide isomerase (PDI)	55	ER	-[Cys-Gly-His-Cys] ₂ -
Ca binding protein 1 (CaBP1)	49	ER	-[Cys-Gly-His-Cys] ₂ -
Ca binding protein 1 (Erp72)	72	ER	-[Cys-Gly-His-Cys] ₃ -
Phospholipase $C\gamma$ (PLC γ)	61	ER	-[Cys-Gly-His-Cys] ₂ -

ER, endoplasmic reticulum; Mt, mitochondria.

Loci Species		Life span	Putative protein function	Reference
[sir-2]	S. cerevisiae	+40%	Histone deacetylase	22
mth	D. melanogaster	+35%*	G protein coupling receptor	35
indy	D. melanogaster	+50%***	Sodium dicarboxylate cotransporter	51
[sod]	D. melanogaster	+48%	Superoxide dismutase	56
[sod/cat]	D. melanogaster	+30%	Superoxide dismutase/catalase	49
[msra]	D. melanogaster	+70%	Methionine sulfoxide reductase	52
age-1	C. elegans	+110%***	PI3K p110 subunit	25
daf-2	C. elegans	+100%*	Insulin-like/IGF-I receptor	29
daf-16	C. elegans	ND	Forkhead transcription factor	34
clk-1	C. elegans	+175%*	Coenzyme Q10 synthesis	32
isp-1	C. elegans	+62%	Electron transport complex III	12
inr	D. melanogaster	+85%	Insulin-like receptor	61
chico	D. melanogaster	+48%**	Insulin receptor substrate	8
prop-1	Mouse	+64%*	Pituitary function	3
pit-1	Mouse	+40%*	Pituitary function	14
p66shc	Mouse	+30%*	Tyrosine receptor adaptor	39
ghr	Mouse	+55%*	Growth hormone receptor	71
inr	Mouse	+18%*	Insulin receptor	2
igf1r	Mouse	+33%*	IGF-1 receptor	19
[trx]	Mouse	+35%**	Thioredoxin	41

TABLE 2. SUMMARY TABLE FOR EXTENDED LIFE SPAN MUTANTS IN EXPERIMENTAL MODEL ORGANISMS

When life span varies among sex, longest life spans are adopted. The listed life spans are average (*), median (**), or maximum (***) depending on the source, and no symbol means not described in the source. Life spans of hetrozygous mutants are indicated in case lethal phenotypes were observed in homozygous mutants. [], transgenic strain or augmented expressions of the loci; ND, not described in detail or ineffective effect by locus alone.

it may reduce ROS, potential by-products of aerobic respiration. Reduction of harmful intermediates may explain why CR treatment can extend life span for most species.

LIFE SPAN EXTENSION BY AUGMENTED EXPRESSION OF TRX IN THE MODEL ANIMAL

In mammals, at least two studies of knockout mice suggested evidence for a direct connection between oxidative stress and life span. One study showed that the p66shc-/- mouse had a significantly extended life span and enhanced resistance to paraquat-induced oxidative stress (39). The ablation of p66shc-/also enhanced cellular resistance to plasma low-density lipoprotein oxidation, arterial oxidation epitopes, and early antherogenic lesions (46). Another study showed that the methionine sulfoxide reductase A (msra) -/- mouse had a shorter life span and high sensitivity toward oxidative stress (42). As for the biological function of msra, it catalyzes the reduction of oxidized methionine in protein by converting methionine sulfoxide to methionine. The catalytic enzyme reaction is completely dependent on the TRX redox system. TRX can serve as an electron donor in order to reduce oxidized form of msra, therefore, the effective reaction of msra is closely associated with TRX, TRX reductase, and NADPH. Meanwhile, Ruan et al. recently reported that msra transgenic animals showed significant resistance to paraquat-induced oxidative stress and extended life span (52). It will be then intriguing to study further possible interactions between msra and TRX on the role of msra-mediated life span extension.

As TRX is a strong antioxidant protein that has significant quenching capability against induced oxidative stress, it serves to reduce cellular oxidative stress significantly (40, 44). Although age-associated changes of the TRX system have not been studied extensively, several reports demonstrate decreased activity along with age (54). It should be also noted that CR treatment can prevent age-associated reduction of TRX as well as TRX reductase in cytoplasm (7). Therefore, the TRX system can be effective in controlling an animal's life span. We have recently produced human TRX transgenic mice driven under the control of the human β -actin promoter to elucidate the direct influence of the TRX redox system on an animal's life span. The TRX transgenic mice showed not only strong resistance to oxiradical stress in a variety of ischemic tissues and organs, but also significant extension of the maximum life span (22%) and median life span (35%) (41, 59) (Fig. 2). These findings demonstrate that antioxidant functions of the cellular redox system, including TRX, effectively contribute to lengthening an animal's life span, even in mammals.

TRX SUPERFAMILY INVOLVING CELLULAR RESPONSES AGAINST OXIDATIVE STRESS

If food availability is limited and cells are under harsh conditions, organisms cannot repair and support their biological activities. As a result, the cellular proliferation is completely blocked at the G_1 cell cycle, and the typical phenotypes of cells will show virtually similar multiple aspects of cellular aging or senescence. Indeed, several genetic mutations have

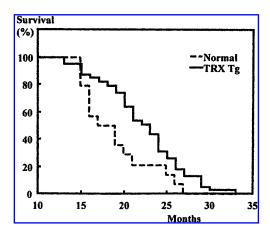


FIG. 2. Life span of the TRX transgenic mice. TRX transgenic mice had significantly extended life span as shown by the survival curves of TRX transgenic (Tg) mice (solid line) and wild-type control (dashed line). Survival curves were plotted according to the Kaplan–Meier method (41).

been identified to show shorter life spans if continuous cellular proliferation is inhibited by DNA damage (11), DNA repair defects (10, 64, 70), enhanced apoptosis (62), or decreased telomere length (17, 66) (Table 3). Those reduced life span mutants are frequently associated with conspicuous growth retardation some time after birth. If the cellular damages exceed a certain point, the cellular antioxidant system, including TRX, is also no longer effective at regulating the redox state and life span. In such cellular conditions, there is accumulating evidence that the expression of TRX is decreasing and the cellular apoptosis pathway is activated. TRX was found to bind and inhibit apoptosis signal regulating kinase (ASK-1), which is an important factor for initiating the p38 mitogen-activated protein (MAP) kinase-mediated apoptosis signal pathway (53). ROS and cytotoxic cytokines including tumor necrosis factor activate ASK-1 through oxidation of TRX to dissociate from ASK-1 (36). In addition, overexpression of

TRX negatively regulates p38 MAP kinase signal transduction and p38 MAP kinase-mediated cytokine production, indicating that TRX has an important role in p38 MAP kinase activation (16).

On the other hand, the expression of TRX tends to be enhanced in tumor cell lines and tissues (27, 38, 45), and transfection of human TRX enhances cellular proliferation (15). At the same time, TRX-2 is uniquely expressed in mitochondria, where it regulates the mitochondrial redox state and plays an important role in cell proliferation (9, 60). TRX-2 was found to form a complex with cytochrome c localized in the mitochondrial matrix, and the releasing of cytochrome c from the mitochondria is significantly enhanced when the expression of TRX-2 is inhibited (48, 60). Overexpression of TRX-2 demonstrates resistance to oxidant-induced apoptosis in human ostosarcoma cells, indicating the critical role in the protection against apoptosis via TRX-2 in mitochondria (6) (Fig. 3). As both TRX and TRX-2 are known as regulators of the manifestation of apoptosis under redox-sensitive caspase (63), potential coordinated actions between TRX and TRX-2 may be possible. However, the function of TRX-1 and TRX-2 does not seem to compensate completely because knockout mice of TRX-2 were found be embryonic lethal (48).

TRX binding protein-2 (TBP-2) identified by yeast two-hybrid screening binds to reduced TRX, but not oxidized TRX (47). TBP-2 was originally identified as a vitamin D upregulated protein 1 (VDUP 1) in HL-60 cells treated with $1\alpha,25$ -dihydroxyvitamin D (5), and is thought to be a negative regulator of TRX (47). Several cysteine residues were identified, and some of them were assumed to bind the active site of TRX (67). Therefore, the overexpression of TBP-2 may block the action of TRX, eventually leading to the inhibition of cellular proliferation. TBP-2 was indeed isolated from a cDNA library enriched for mRNA species that immediately increase by administration of BrdU as a senescence-associated gene (57). Enhanced expression of TBP-2/VDUP 1 was also reported to be sensitive to paraquat-induced oxidative stress (24). On the other hand, a significant reduction in TBP-2 expression was frequently observed in several tumor cell lines and tumor tis-

TABLE 3. SUMMARY TABLE FOR REDUCED LIFE SPAN MUTANTS IN EXPERIMENTAL MODEL ORGANISMS

Loci	Species	Life span	Putative protein function	Reference
sir-2	S. cerevisiae	-50%*	NAD-dependent deacetylase	26
gas-1	C. elegans	-25%	Electron transport complex I	28
mev-1	C. elegans	-30%*	Electron transport complex II	23
top3β	Mouse	-35%*	DNA topoisomeraseβ	31
ttd	Mouse	-50%*	Nucleotide excision repair	10
[<i>p53</i>]	Mouse	-65%***	Tumor suppressor	62
msra	Mouse	-40%*	Methionine sulfoxide reductase	42
ku86	Mouse	-61%*	Double-strand break DNA repair	64
terc	Mouse	ND	Telomerase	17
klotho	Mouse	-92%*	Calcium regulation?	30
wrn	Mouse	(-83%)	DNA helicase	37
lmna	Mouse	-95%	A-type lamins	43

Most reduced life span mutants in lower organisms were omitted. When life span varies among sex, longest life spans are adopted. The listed life spans are average (*), median (**), or maximum (***) depending on the source, and no symbol means not described in the source. Life span of *wrn* is relative to p53 null background. Life spans of hetrozygous mutants are indicated in case lethal phenotypes are observed in homozygous mutants. [], transgenic strain or augmented expressions of the loci; ND, not described in detail or ineffective effect by locus alone.

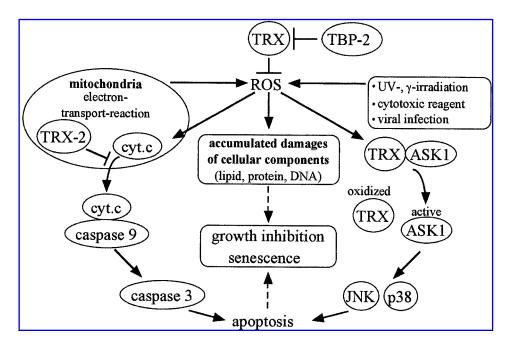


FIG. 3. Multiple functions of TRX and TRX-2 in oxidative stress. TRX and TRX-2 are involved in a variety of signal transductions including ROS-mediated cellular damage and apoptosis. The accumulated damages of cellular components eventually lead to growth inhibition and cellular senescence.

sues, including human primary breast and colon tumors (4, 68). Moreover, a strong antioncogenic histone deacetylase inhibitor, suberoylanilide hydroxamic acid, up-regulates the expression of TBP-2 accompanied by a significant reduction in TRX (4). These reports support the idea that the expression of TRX and TBP-2 are complementarily reflected in cellular proliferation and cellular senescence in vivo and in vitro.

CONCLUDING REMARKS

In conclusion, TRX is a strong redox-regulating antioxidant protein that has significant quenching capability against induced oxidative stress. Accumulating evidence demonstrates that TRX functions efficiently as an antioxidant against deterioration along with the aging process (senescence) via the cellular redox state, and can extend an animal's life span. The TRX system, including TBP-2, can also serve as an index marker for cellular proliferation and senescence based on their correlative expressions with cellular conditions.

ACKNOWLEDGMENTS

We thank Drs. Y. Nishinaka and A. Mitsui for help in preparing the manuscript and valuable discussion.

ABBREVIATIONS

ASK-1, apoptosis signal-regulating kinase; CR, caloric restriction; MAP, mitogen-activated protein; PI3K, phosphatidylinositol 3-kinase; ROS, reactive oxygen species; SOD, super-

oxide dismutase; TBP-2, thioredoxin binding protein-2; TRX, thioredoxin; VDUP 1, vitamin D up-regulated protein 1.

REFERENCES

- 1. Arking R. *Biology of Aging*, 2nd edit. Sunderland, MA: Sinauer Associates, 1998, pp. 311–364.
- Bluher M, Kahn BB, and Kahn CR. Extended longevity in mice lacking the insulin receptor in adipose tissue. *Science* 299: 572–574, 2003.
- 3. Brown-Borg HM, Borg KE, Meliska CJ, and Bartke A. Dwarf mice and the ageing process. *Nature* 384: 33, 1996.
- Butler LM, Zhou X, Xu WS, Scher HI, Rifkind RA, Marks PA, and Richon VM. The histone deacetylase inhibitor SAHA arrests cancer cell growth, up-regulates thioredoxinbinding protein-2, and down-regulates thioredoxin. *Proc* Natl Acad Sci U SA 99: 11700–11705, 2002.
- Chen KS and DeLuca HF. Isolation and characterization of a novel cDNA from HL-60 cells treated with 1,25-dihydroxy vitamin D-3. *Biochim Biophys Acta* 1219: 26–32, 1994.
- Chen Y, Cai J, Murphy TJ, and Jones DP. Overexpressed human mitochondrial thioredoxin confers resistance to oxidant-induced apoptosis in human osteosarcoma cells. *J Biol Chem* 277: 33242–33248, 2002.
- 7. Cho CG, Kim HJ, Chung SW, Jung KJ, Shim KH, Yu BP, Yodoi J, and Chung HY. Modulation of glutathione and thioredoxin systems by calorie restriction during the aging process. *Exp Gerontol* 38: 539–548, 2003.
- 8. Clancy DJ, Gems D, Harshman LG, Oldham S, Stocker H, Hafen E, Leevers SJ, and Partridge L. Extension of lifespan by loss of CHICO, a *Drosophila* insulin receptor substrate protein. *Science* 292: 104–106, 2001.

 Damdimopoulos AE, Miranda-Vizuete A, Pelto-Huikko M, Gustafsson JA, and Spyrou G. Human mitochondrial thioredoxin. Involvement in mitochondrial membrane potential and cell death. *J Biol Chem* 277: 33249–33257, 2002.

- 10. De Boer J, Andressoo JO, De Wit J, Huijmans J, Beems RB, van Steeg H, Weeda G, Van Der Horst GT, van Leeuwen W, Themmen AP, Meradji M, and Hoeijmakers JH. Premature aging in mice deficient in DNA repair and transcription. *Science* 296: 1276–1279, 2002.
- Dolle ME, Giese H, Hopkins CL, Martus HJ, Hausdorff JM, and Vijg J. Rapid accumulation of genome rearrangements in liver but not in brain of old mice. *Nat Genet* 17: 431–434, 1997.
- 12. Feng J, Bussiere F, and Hekimi S. Mitochondrial electron transport is a key determinant of life span in *Caenorhabditis elegans*. *Dev Cell* 1: 633–644, 2001.
- Finch CE. Longevity, Senescence, and the Genome. Chicago, IL: University of Chicago Press, 1990, pp. 497
 –566.
- 14. Flurkey K, Papaconstantinou J, Miller RA, and Harrison DE. Lifespan extension and delayed immune and collagen aging in mutant mice with defects in growth hormone production. *Proc Natl Acad Sci U S A* 98: 6736–6741, 2001.
- 15. Gallegos A, Gasdaska JR, Taylor CW, Paine-Murrieta GD, Goodman D, Gasdaska PY, Berggren M, Briehl MM, and Powis G. Transfection with human thioredoxin increases cell proliferation and a dominant-negative mutant thioredoxin reverses the transformed phenotype of human breast cancer cells. *Cancer Res* 56: 5765–5770, 1996.
- 16. Hashimoto S, Matsumoto K, Gon Y, Furuichi S, Maruoka S, Takeshita I, Hirota K, Yodoi J, and Horie T. Thioredoxin negatively regulates p38 MAP kinase activation and IL-6 production by tumor necrosis factor-alpha. *Biochem Biophys Res Commun* 258: 443–447, 1999.
- Herrera E, Samper E, Martin-Caballero J, Flores JM, Lee HW, and Blasco MA. Disease states associated with telomerase deficiency appear earlier in mice with short telomeres. *EMBO J* 18: 2950–2960, 1999.
- Holmgren A. Thioredoxin. *Annu Rev Biochem* 54: 237–271, 1985.
- Holzenberger M, Dupont J, Ducos B, Leneuve P, Geloen A, Even PC, Cervera P, and Le Bouc Y. IGF-1 receptor regulates lifespan and resistance to oxidative stress in mice. *Nature* 421: 182–187, 2003.
- 20. Honda Y and Honda S. The daf-2 gene network for longevity regulates oxidative stress resistance and Mnsuperoxide dismutase gene expression in *Caenorhabditis elegans*. *FASEB J* 13: 1385–1393, 1999.
- 21. Honda Y and Honda S. Oxidative stress and life span determination in the nematode *Caenorhabditis elegans*. *Ann NYAcad Sci* 959: 466–474, 2002.
- Imai S, Armstrong CM, Kaeberlein M, and Guarente L. Transcriptional silencing and longevity protein Sir2 is an NAD-dependent histone deacetylase. *Nature* 403: 795–800, 2000.
- 23. Ishii N, Fujii M, Hartman PS, Tsuda M, Yasuda K, Senoo-Matsuda N, Yanase S, Ayusawa D, and Suzuki K. A mutation in succinate dehydrogenase cytochrome b causes oxidative stress and ageing in nematodes. Nature 394: 694–697, 1998.

24. Joguchi A, Otsuka I, Minagawa S, Suzuki T, Fujii M, and Ayusawa D. Overexpression of VDUP1 mRNA sensitizes HeLa cells to paraquat. *Biochem Biophys Res Commun* 293: 293–297, 2002.

- Johnson TE. Increased life-span of age-1 mutants in Caenorhabditis elegans and lower Gompertz rate of aging. Science 249: 908–912, 1990.
- Kaeberlein M, McVey M, and Guarente L. The SIR2/3/4 complex and SIR2 alone promote longevity in Saccharomyces cerevisiae by two different mechanisms. Genes Dev 13:2570–2580, 1999.
- 27. Kahlos K, Soini Y, Saily M, Koistinen P, Kakko S, Paakko P, Holmgren A, and Kinnula VL. Up-regulation of thioredoxin and thioredoxin reductase in human malignant pleural mesothelioma. *Int J Cancer* 95: 198–204, 2001.
- 28. Kayser EB, Morgan PG, Hoppel CL, and Sedensky MM. Mitochondrial expression and function of GAS-1 in *Caenorhabditis elegans. J Biol Chem* 276: 20551–20558, 2001.
- 29. Kimura KD, Tissenbaum HA, Liu Y, and Ruvkun G. daf-2, an insulin receptor-like gene that regulates longevity and diapause in *Caenorhabditis elegans*. *Science* 277: 942–946, 1997.
- 30. Kuro-o M, Matsumura Y, Aizawa H, Kawaguchi H, Suga T, Utsugi T, Ohyama Y, Kurabayashi M, Kaname T, Kume E, Iwasaki H, Iida A, Shiraki-Iida T, Nishikawa S, Nagai R, and Nabeshima YI. Mutation of the mouse klotho gene leads to a syndrome resembling ageing. *Nature* 390: 45–51, 1997.
- 31. Kwan KY and Wang JC. Mice lacking DNA topoisomerase IIIbeta develop to maturity but show a reduced mean lifespan. *Proc Natl Acad Sci U S A* 98: 5717–5721, 2001.
- 32. Lakowski B and Hekimi S. Determination of life-span in *Caenorhabditis elegans* by four clock genes. *Science* 272: 1010–1013, 1996.
- 33. Larsen PL. Aging and resistance to oxidative damage in *Caenorhabditis elegans. Proc Natl Acad Sci U S A* 90: 8905–8909, 1993.
- 34. Lin K, Dorman JB, Rodan A, and Kenyon C. daf-16: an HNF-3/forkhead family member that can function to double the life-span of *Caenorhabditis elegans*. *Science* 278: 1319–1322, 1997.
- 35. Lin YJ, Seroude L, and Benzer S. Extended life-span and stress resistance in the *Drosophila* mutant methuselah. *Science* 282: 943–946, 1998.
- Liu H, Nishitoh H, Ichijo H, and Kyriakis JM. Activation of apoptosis signal-regulating kinase 1 (ASK1) by tumor necrosis factor receptor-associated factor 2 requires prior dissociation of the ASK1 inhibitor thioredoxin. *Mol Cell Biol* 20: 2198–2208, 2000.
- 37. Lombard DB, Beard C, Johnson B, Marciniak RA, Dausman J, Bronson R, Buhlmann JE, Lipman R, Curry R, Sharpe A, Jaenisch R, and Guarente L. Mutations in the WRN gene in mice accelerate mortality in a p53-null background. *Mol Cell Biol* 20: 3286–3291, 2000.
- 38. Matsutani Y, Yamauchi A, Takahashi R, Ueno M, Yoshikawa K, Honda K, Nakamura H, Kato H, Kodama H, Inamoto T, Yodoi J, and Yamaoka Y. Inverse correlation of thioredoxin expression with estrogen receptor- and p53-

- dependent tumor growth in breast cancer tissues. *Clin Cancer Res* 7: 3430–3436, 2001.
- 39. Migliaccio E, Giorgio M, Mele S, Pelicci G, Reboldi P, Pandolfi PP, Lanfrancone L, and Pelicci PG. The p66shc adaptor protein controls oxidative stress response and life span in mammals. *Nature* 402: 309–313, 1999.
- Mitsui A, Hirakawa T, and Yodoi J. Reactive oxygenreducing and protein-refolding activities of adult T cell leukemia-derived factor/human thioredoxin. *Biochem Bio*phys Res Commun 186: 1220–1226, 1992.
- 41. Mitsui A, Hamuro J, Nakamura H, Kondo N, Hirabayashi Y, Ishizaki-Koizumi S, Hirakawa T, Inoue T, and Yodoi J. Overexpression of human thioredoxin in transgenic mice controls oxidative stress and life span. *Antioxid Redox Signal* 4: 693–696, 2002.
- 42. Moskovitz J, Bar-Noy S, Williams WM, Requena J, Berlett BS, and Stadtman ER. Methionine sulfoxide reductase (MsrA) is a regulator of antioxidant defense and lifespan in mammals. *Proc Natl Acad Sci U S A* 98: 12920–12925, 2001.
- 43. Mounkes LC, Kozlov S, Hernandez L, Sullivan T, and Stewart CL. A progeroid syndrome in mice is caused by defects in A-type lamins. *Nature* 423: 298–301, 2003.
- Nakamura H, Nakamura K, and Yodoi J. Redox regulation of cellular activation. *Annu Rev Immunol* 15: 351–369, 1997.
- 45. Nakamura H, Bai J, Nishinaka Y, Ueda S, Sasada T, Ohshio G, Imamura M, Takabayashi A, Yamaoka Y, and Yodoi J. Expression of thioredoxin and glutaredoxin, redox-regulating proteins, in pancreatic cancer. *Cancer Detect Prev* 24: 53–60, 2000.
- 46. Napoli C, Martin-Padura I, de Nigris F, Giorgio M, Mansueto G, Somma P, Condorelli M, Sica G, De Rosa G, and Pelicci P. Deletion of the p66Shc longevity gene reduces systemic and tissue oxidative stress, vascular cell apoptosis, and early atherogenesis in mice fed a high-fat diet. *Proc Natl Acad Sci U S A* 100: 2112–2116, 2003.
- 47. Nishiyama A, Matsui M, Iwata S, Hirota K, Masutani H, Nakamura H, Takagi Y, Sono H, Gon Y, and Yodoi J. Identification of thioredoxin-binding protein-2/vitamin D(3) upregulated protein 1 as a negative regulator of thioredoxin function and expression. *J Biol Chem* 274: 21645–21650, 1999.
- 48. Nonn L, Williams RR, Erickson RP, and Powis G. The absence of mitochondrial thioredoxin 2 causes massive apoptosis, exencephaly, and early embryonic lethality in homozygous mice. *Mol Cell Biol* 23: 916–922, 2003.
- 49. Orr WC and Sohal RS. Extension of life-span by overexpression of superoxide dismutase and catalase in *Drosophila melanogaster Science* 263: 1128–1130, 1994.
- 50. Parkes TL, Elia AJ, Dickinson D, Hilliker AJ, Phillips JP, and Boulianne GL. Extension of *Drosophila* lifespan by overexpression of human SOD1 in motorneurons. *Nat Genet* 19: 171–174, 1998.
- 51. Rogina B, Reenan RA, Nilsen SP, and Helfand SL. Extended life-span conferred by cotransporter gene mutations in *Drosophila*. *Science* 290: 2137–2140, 2000.
- 52. Ruan H, Tang XD, Chen ML, Joiner ML, Sun G, Brot N, Weissbach H, Heinemann SH, Iverson L, Wu CF, Hoshi T, Chen ML, Joiner MA, and Heinemann SH. High-quality life extension by the enzyme peptide methionine sulfoxide reductase. *Proc Natl Acad Sci U S A* 99: 2748–2753, 2002.

- 53. Saitoh M, Nishitoh H, Fujii M, Takeda K, Tobiume K, Sawada Y, Kawabata M, Miyazono K, and Ichijo H. Mammalian thioredoxin is a direct inhibitor of apoptosis signalregulating kinase (ASK) 1. EMBO J 17: 2596–2606, 1998.
- 54. Santa Maria C and Machado A. Effects of development and ageing on pulmonary NADPH-cytochrome c reductase, glutathione peroxidase, glutathione reductase and thioredoxin reductase activities in male and female rats. Mech Ageing Dev 37: 183–195, 1986.
- Scott BA, Avidan MS, Crowder CM. Regulation of hypoxic death in *C. elegans* by the insulin/IGF receptor homolog DAF-2. *Science* 296: 2388–2391, 2002.
- Sun J, Folk D, Bradley TJ, and Tower J. Induced overexpression of mitochondrial Mn-superoxide dismutase extends the life span of adult *Drosophila melanogaster Ge*netics 161: 661–672, 2002.
- Suzuki T, Minagawa S, Michishita E, Ogino H, Fujii M, Mitsui Y, and Ayusawa D. Induction of senescence-associated genes by 5-bromodeoxyuridine in HeLa cells. *Exp Gerontol* 36: 465–474, 2001.
- 58. Tagaya Y, Maeda Y, Mitsui A, Kondo N, Matsui H, Hamuro J, Brown N, Arai K, Yokota T, Wakasugi H, and Yodoi J. ATL-derived factor (ADF), an IL-2 receptor/Tac inducer homologous to thioredoxin; possible involvement of dithiol-reduction in the IL-2 receptor induction. *EMBO J* 8: 757–764, 1989.
- Takagi Y, Mitsui A, Nishiyama A, Nozaki K, Sono H, Gon Y, Hashimoto N, and Yodoi J. Overexpression of thioredoxin in transgenic mice attenuates focal ischemic brain damage. *Proc Natl Acad Sci U S A* 96: 4131–4136, 1999.
- 60. Tanaka T, Hosoi F, Yamaguchi-Iwai Y, Nakamura H, Masutani H, Ueda S, Nishiyama A, Takeda S, Wada H, Spyrou G, and Yodoi J. Thioredoxin-2(TRX-2) is an essential gene regulating mitochondria-dependent apoptosis. *EMBO J* 21: 1695–1703, 2002.
- 61. Tatar M, Kopelman A, Epstein D, Tu MP, Yin CM, and Garofalo RS. A mutant *Drosophila* insulin receptor homolog that extends life-span and impairs neuroendocrine function. *Science* 292: 107–110, 2001.
- 62. Tyner SD, Venkatachalam S, Choi J, Jones S, Ghebranious N, Igelmann H, Lu X, Soron G, Cooper B, Brayton C, Hee Park S, Thompson T, Karsenty G, Bradley A, and Donehower LA. p53 mutant mice that display early ageing-associated phenotypes. *Nature* 415: 45–53, 2002.
- Ueda S, Masutani H, Nakamura H, Tanaka T, Ueno M, and Yodoi J. Redox control of cell death. *Antioxid Redox Signal* 4: 405–414, 2002.
- Vogel H, Lim DS, Karsenty G, Finegold M, and Hasty P. Deletion of Ku86 causes early onset of senescence in mice. *Proc Natl Acad Sci U S A* 96: 10770–10775, 1999.
- 65. Wakasugi H, Rimsky L, Mahe Y, Kamel AM, Fradelizi D, Tursz T, and Bertoglio J. Epstein–Barr virus-containing B-cell line produces an interleukin 1 that it uses as a growth factor. *Proc Natl Acad Sci U S A* 84: 804–808, 1987.
- 66. Wong KK, Maser RS, Bachoo RM, Menon J, Carrasco DR, Gu Y, Alt FW, and DePinho RA. Telomere dysfunction and Atm deficiency compromises organ homeostasis and accelerates ageing. *Nature* 421: 643–648, 2003.
- 67. Yamanaka H, Maehira F, Oshiro M, Asato T, Yanagawa Y, Takei H, and Nakashima Y. A possible interaction of

thioredoxin with VDUP1 in HeLa cells detected in a yeast two-hybrid system. *Biochem Biophys Res Commun* 271: 796–800, 2000.

- Yang X, Young LH, and Voigt JM. Expression of a vitamin D-regulated gene (VDUP-1) in untreated- and MNU-treated rat mammary tissue. *Breast Cancer Res Treat* 48: 33–44, 1998.
- Yodoi J and Uchiyama T. Diseases associated with HTLV-I: virus, IL-2 receptor dysregulation and redox regulation. *Immunol Today* 13: 405–411, 1992.
- Yu CE, Oshima J, Fu YH, Wijsman EM, Hisama F, Alisch R, Matthews S, Nakura J, Miki T, Ouais S, Martin GM, Mulligan J, and Schellenberg GD. Positional cloning of the Werner's syndrome gene. *Science* 272: 258–262, 1996.
- Zhou Y, Xu BC, Maheshwari HG, He L, Reed M, Lozykowski M, Okada S, Cataldo L, Coschigamo K, Wagner TE, Baumann G, and Kopchick JJ. A mammalian

model for Laron syndrome produced by targeted disruption of the mouse growth hormone receptor/binding protein gene (the Laron mouse). *Proc Natl Acad Sci U S A* 94: 13215–13220, 1997.

Address reprint requests to:
Junji Yodoi, M.D., Ph.D.
Department of Biological Responses
Institute for Virus Research, Kyoto University
53 Shogoin
Kawahara-cho
Sakyo-ku
Kyoto, 606-8507, Japan

E-mail: yodoi@virus1.virus.kyoto-u.æ.jp

Received for publication April 28, 2003; accepted July 14, 2003.

This article has been cited by:

- 1. Tatsuya Oba, Ryosuke Tatsunami, Keisuke Sato, Kyohei Takahashi, Zhihui Hao, Yoshiko Tampo. 2012. Methylglyoxal has deleterious effects on thioredoxin in human aortic endothelial cells. *Environmental Toxicology and Pharmacology* **34**:2, 117-126. [CrossRef]
- Léonie G.M. Boender, Antonius J.A. Maris, Erik A.F. Hulster, Marinka J.H. Almering, Ida J. Klei, Marten Veenhuis, Johannes H. Winde, Jack T. Pronk, Pascale Daran-Lapujade. 2011. Cellular responses of Saccharomyces cerevisiae at near-zero growth rates: transcriptome analysis of anaerobic retentostat cultures. FEMS Yeast Research 11:8, 603-620. [CrossRef]
- 3. Yi Liu , Yanzhuo Ma , Rutao Wang , Chenhai Xia , Rongqing Zhang , Kun Lian , Ronghua Luan , Lu Sun , Lu Yang , Wayne B. Lau , Haichang Wang , Ling Tao . 2011. Advanced Glycation End Products Accelerate Ischemia/Reperfusion Injury Through Receptor of Advanced End Product/ Nitrative Thioredoxin Inactivation in Cardiac Microvascular Endothelial Cells. *Antioxidants & Redox Signaling* 15:7, 1769-1778. [Abstract] [Full Text HTML] [Full Text PDF] [Full Text PDF] with Links]
- 4. Baruch E. Bulvik, Eduard Berenshtein, Abraham Marim Konijn, Leonid Grinberg, Vladimir Vinokur, Ron Eliashar, Mordechai Chevion. 2011. Aging is an organ-specific process: changes in homeostasis of iron and redox proteins in the rat. *AGE*. [CrossRef]
- 5. Meili Li, Lin Wang, Xiaoming Ren, Chunfu Zheng. 2011. Host cell targets of tegument protein VP22 of herpes simplex virus 1. *Archives of Virology* **156**:6, 1079-1084. [CrossRef]
- 6. Lizette Gil del Valle. 2011. Oxidative stress in aging: Theoretical outcomes and clinical evidences in humans. *Biomedicine & Aging Pathology* 1:1, 1-7. [CrossRef]
- 7. Michael Lustgarten, Florian L. Muller, Holly Van RemmenAn Objective Appraisal of the Free Radical Theory of Aging 177-202. [CrossRef]
- 8. Lizette Gil del Valle. 2010. Oxidative stress in aging: Theoretical outcomes and clinical evidences in humans. *Biomedicine & Pharmacotherapy*. [CrossRef]
- 9. Y. Kamimoto, T. Sugiyama, T. Kihira, L. Zhang, N. Murabayashi, T. Umekawa, K. Nagao, N. Ma, N. Toyoda, J. Yodoi, N. Sagawa. 2010. Transgenic mice overproducing human thioredoxin-1, an antioxidative and anti-apoptotic protein, prevents diabetic embryopathy. *Diabetologia* **53**:9, 2046-2055. [CrossRef]
- 10. Hiroaki Okuyama, Toru Yoshida, Aoi Son, Shin-ichi Oka, Dongmei Wang, Rika Nakayama, Hiroshi Masutani, Hajime Nakamura, Yo-ichi Nabeshima, Junji Yodoi. 2009. Thioredoxin Binding Protein 2 Modulates Natural Killer T Cell-Dependent Innate Immunity in the Liver: Possible Link to Lipid Metabolism. Antioxidants & Redox Signaling 11:10, 2585-2593. [Abstract] [Full Text HTML] [Full Text PDF] [Full Text PDF with Links]
- 11. R. Luan, S. Liu, T. Yin, W. B. Lau, Q. Wang, W. Guo, H. Wang, L. Tao. 2009. High glucose sensitizes adult cardiomyocytes to ischaemia/reperfusion injury through nitrative thioredoxin inactivation. *Cardiovascular Research* 83:2, 294-302. [CrossRef]
- 12. Rui Bao, Yaru Zhang, Cong-Zhao Zhou, Yuxing Chen. 2009. Structural and mechanistic analyses of yeast mitochondrial thioredoxin Trx3 reveal putative function of its additional cysteine residues. *Biochimica et Biophysica Acta (BBA) Proteins and Proteomics* **1794**:4, 716-721. [CrossRef]
- 13. Vladimir Vinokur, Leonid Grinberg, Eduard Berenshtein, Menachem Gross, Jackob Moskovitz, Abraham Z. Reznick, Mordechai Chevion, Ron Eliashar. 2009. Methionine-centered redox cycle in organs of the aero-digestive tract of young and old rats. *Biogerontology* 10:1, 43-52. [CrossRef]
- 14. Jamie Case, David A. Ingram, Laura S. Haneline. 2008. Oxidative Stress Impairs Endothelial Progenitor Cell Function. *Antioxidants & Redox Signaling* **10**:11, 1895-1907. [Abstract] [Full Text HTML] [Full Text PDF] [Full Text PDF with Links]
- 15. Yi-Ling Huang, Chun-Yu Chuang, Fung-Chang Sung, Chia-Yang Chen. 2008. Thioredoxin Overexpression Modulates Remodeling Factors in Stress Responses to Cigarette Smoke. *Journal of Toxicology and Environmental Health, Part A* **71**:22, 1490-1498. [CrossRef]

- 16. Malcolm J. Jackson. 2008. Redox regulation of skeletal muscle. IUBMB Life 60:8, 497-501. [CrossRef]
- 17. Yosuke Funato, Hiroaki Miki. 2007. Nucleoredoxin, a Novel Thioredoxin Family Member Involved in Cell Growth and Differentiation. *Antioxidants & Redox Signaling* **9**:8, 1035-1058. [Abstract] [Full Text PDF] [Full Text PDF with Links]
- 18. Hangxiang Zhang, Ling Tao, Xiangying Jiao, Erhe Gao, Bernard L. Lopez, Theodore A. Christopher, Walter Koch, Xin L. Ma. 2007. Nitrative thioredoxin inactivation as a cause of enhanced myocardial ischemia/reperfusion injury in the aging heart. *Free Radical Biology and Medicine* **43**:1, 39-47. [CrossRef]
- 19. Su-Jung Kim, Eui-Man Jung, Hyun-Joo Jung, Yun Seon Song, Eun-Hee Park, Chang-Jin Lim. 2007. Cellular functions and transcriptional regulation of a third thioredoxin from Schizosaccharomyces pombe. *Canadian Journal of Microbiology* **53**:6, 775-783. [CrossRef]
- 20. Christopher Horst Lillig, Arne Holmgren. 2007. Thioredoxin and Related Molecules–From Biology to Health and Disease. *Antioxidants & Redox Signaling* **9**:1, 25-47. [Abstract] [Full Text PDF] [Full Text PDF with Links]
- 21. Eun Kyung Go, Kyung Jin Jung, Ji Min Kim, Hyunae Lim, Hyeang Kwan Lim, Byung Pal Yu, Hae Young Chung. 2007. Betaine Modulates Age-Related NF-#B by Thiol-Enhancing Action. *Biological & Pharmaceutical Bulletin* **30**:12, 2244-2249. [CrossRef]
- 22. Pamela Maher . 2006. Redox Control of Neural Function: Background, Mechanisms, and Significance. *Antioxidants & Redox Signaling* **8**:11-12, 1941-1970. [Abstract] [Full Text PDF] [Full Text PDF with Links]
- 23. Evgenia Makrantonaki, James Adjaye, Ralf Herwig, Thore C. Brink, Detlef Groth, Claus Hultschig, Hans Lehrach, Christos C. Zouboulis. 2006. Age-specific hormonal decline is accompanied by transcriptional changes in human sebocytes in vitro. *Aging Cell* 5:4, 331-344. [CrossRef]
- 24. H. Fai Poon, Vittorio Calabrese, Menotti Calvani, Professor D. Allan Butterfield. 2006. Proteomics Analyses of Specific Protein Oxidation and Protein Expression in Aged Rat Brain and Its Modulation by L-Acetylcarnitine: Insights Into the Mechanisms of Action of This Proposed Therapeutic Agent for CNS Disorders Associated with Oxidative Stress. *Antioxidants & Redox Signaling* 8:3-4, 381-394. [Abstract] [Full Text PDF] [Full Text PDF] with Links]
- 25. Graham D. Jack, E. Andrew Mead, James F. Garst, M. Carla Cabrera, Andrea M. DeSantis, Stephen M. Slaughter, Jody Jervis, Andrew I. Brooks, Malcolm Potts, Richard F. Helm. 2006. Long term metabolic arrest and recovery of HEK293 spheroids involves NF-#B signaling and sustained JNK activation. *Journal of Cellular Physiology* 206:2, 526-536. [CrossRef]
- 26. Kumuda C. Das . 2005. Thioredoxin and Its Role in Premature Newborn Biology. *Antioxidants & Redox Signaling* **7**:11-12, 1740-1743. [Abstract] [Full Text PDF] [Full Text PDF] with Links]
- 27. Richard G. Cutler . 2005. Oxidative Stress and Aging: Catalase Is a Longevity Determinant Enzyme. *Rejuvenation Research* **8**:3, 138-140. [Abstract] [Full Text PDF] [Full Text PDF with Links]
- 28. Anne Burke-Gaffney, Matthew E.J. Callister, Hajime Nakamura. 2005. Thioredoxin: friend or foe in human disease?. *Trends in Pharmacological Sciences* **26**:8, 398-404. [CrossRef]
- 29. Hajime Nakamura . 2005. Thioredoxin and Its Related Molecules: Update 2005. *Antioxidants & Redox Signaling* **7**:5-6, 823-828. [Abstract] [Full Text PDF] [Full Text PDF with Links]
- 30. Hiroyuki Kakimaru, Hiroko Kataoka, Koh-ichi Enomoto, Nobuyuki Kumahashi, Akihiro Ohira, Junji Yodoi, Yuji Uchio, Mitsuo Ochi. 2005. Thioredoxin Gene Expression in Rat Knee Articular Cartilage After Full-Thickness Injury. *Connective Tissue Research* **46**:1, 27-32. [CrossRef]
- 31. Isabelle Petropoulos, Bertrand Friguet. 2005. Protein maintenance in aging and replicative senescence: a role for the peptide methionine sulfoxide reductases. *Biochimica et Biophysica Acta (BBA) Proteins and Proteomics* **1703**:2, 261-266. [CrossRef]
- 32. Li Chen, Donald C. Rio, Gabriel G. Haddad, Enbo Ma. 2004. Regulatory role of dADAR in ROS metabolism in Drosophila CNS. *Molecular Brain Research* **131**:1-2, 93-100. [CrossRef]

33. Tory M. Hagen . 2003. Oxidative Stress, Redox Imbalance, and the Aging Process. *Antioxidants & Redox Signaling* **5**:5, 503-506. [Citation] [Full Text PDF] [Full Text PDF with Links]